

A Revolution By Probiotic And Paraprobiotic In Food Industry: Review

¹Salunke Ramkrishna, ²Chelladurai Chellamboli and ³Devkotte Anupama

¹MIT College of Food Technology, MIT College of Food Technology, Loni Kalbhor, Pune-412201 (India)

²Assistant Professor, Department of Ethical Science and Food Technology, MIT College of Food Technology, MIT Art Design and Technology University, Loni Kalbhor, Pune-412201 (India)

³Associate Professor & Head, Department of Ethical Science and Food Technology, MIT College of Food Technology, MIT Art Design and Technology University, Loni Kalbhor, Pune-412201 (India)

ARTICLE DETAILS

Article History

Published Online: 16 Sep 2019

Keywords

Probiotics, Paraprobiotics, Human Health

*Corresponding Author

Email: anu_devkotte[at]yahoo.com

ABSTRACT

This review article focuses on the bio-food revolution arising by use of probiotics and paraprobiotic products for eliminating the deficiency and improve the human immune system to healthier state. Paraprobiotic products aid in improving the digestive system, reducing mental and nervous illnesses in humans. Probiotics and Paraprobiotics can be used as medicine which can inhibit the activation of unwanted organisms in a human system and prevent the diseases. In the production of probiotics and paraprobiotic products, live or inactivated microorganisms are used. Day to day humans come across extensive metabolic activities for which such organisms are needed by human body. In human body, microorganisms play vital role, which may be beneficial or harmful. In Food Industry the development will concentrate on enhancement of beneficial organisms and contain harmful one to avoid risk factors to human body. Therefore, the focus area of this review paper is to study beneficial microorganisms and investigated same for the manufacturing of paraprobiotic products. This study aims at functionality and scope of probiotics and Paraprobiotics in future in this area taking cognizance of past work. This review concludes that there is significant impact of probiotics and paraprobiotics products on human health. Same can be used as bio-food for eradicating imminent deficiency issues in human body.

1. Introduction

In the human body, there are numerous varieties of beneficial microorganisms are present in an external and internal area which includes skin, teeth, hand, stomach, intestine, mouth and also clothes. Even in the human gastrointestinal tract (GIT), these microorganism plays a very prominent role in digestion and also maintaining good health of human being. The human body is a host of microorganism. Those microorganisms present in the human body are mostly useful for functional activity of human beings.

Probiotics is a Greek word "Pro-Bios" means for life. This term first introduced by Kollath in 1953 and used by Lilly and Stillwell in 1965. The first probiotic product was fermented milk used for human consumption. At the time of 20th century, Russian scientist "Elie Metchnikoff" (Nobel Prize winner) who suggested the long healthy life of Bulgarian people after consuming the fermented milk product which decreased the toxic microbial activity (Arvind Kumar and S. Shafat, 2010). After that in the 1960s dairy industry was beginning to promote fermented milk and milk products contain *Lactobacillus acidophilus*. Commonly probiotics are defined as "live microorganism is orally taken in adequate amount to maintain good health or disease free". It is reported that health benefits are provided due to interaction between the gastrointestinal microorganisms and immune system. Probiotic health benefits depend on the dose taken by the humans and support the immune system of the human body (Caroline N. de Almada. et

al., 2016). In general, good health is provided by probiotic foods. Probiotics are classified into three categories according to the site and action of microorganisms they are, 1) Direct action on GIT microbes, 2) Direct action on epithelium and mucous layer, 3) Action on the immune system and other organs like liver, brain etc., (Rijker et al., 2010).

Probiotics are used to control diarrhea, lactose intolerance (Li et al., 2012), irritable bowel syndrome (Ducrotte et al., 2012), some type of cancer, insulin resistance (Hsieh et al., 2013), cholesterol (Bordoni et al., 2013), blood pressure (Chiang and Pan, 2012) and obesity (Arora et al., 2013). Different types of microorganisms like bacteria and yeasts which contribute the probiotic properties are used in food and clinical treatment (Amara and Shilb, 2013). In the production of probiotics and paraprobiotics mostly microbes are used which is given in Table 1. Some species are traditionally used in the production of food and beverages like fermented milk, yogurts, idli, chees, etc. In orange juice and dehydrated food contain stressful microorganism due to their low pH and water activity (Anekella and Orsat, 2013; Betoret et al., 2003; Krasaekoopt et al., 2014). At the time of production of probiotics, microorganisms are supplement after thermal treatment to avoid the chance of microbial contamination (Rokka and Rantamaki., 2010). Although, now research is focused on inactivation of probiotic microorganisms are known as paraprobiotics.

Table-1
Application of Various Probiotic organisms

Sr. No.	Organisms	Product/ used in/ find in	References
1	<i>Lactobacillus acidophilus</i>	Fruit- based ice cream, Lactic acid production	Senanayake et al., 2013; Rossana liguori, 2015
2	<i>Lactobacillus sporogenes</i>	Egg production and quality of yolk cholesterol	Panda et al., 2008
3	<i>Lactobacillus plantarum</i>	starter culture in caper berry fermentation	Palomino et al., 2015
4	<i>Lactobacillus rhamnosus</i>	Naturally, in intestines prevent the growth of harmful bacteria	-
5	<i>Lactobacillus delbrueckii</i>	Bread	Plessas et al., 2008
6	<i>Lactobacillus reuteri</i>	Inhibit Some Food-borne Pathogens in UF-Feta-Cheese	Mohamadi Sani et al., 2005
7	<i>Lactobacillus fermentum</i>	Manufacture of traditional Iberian dry-fermented sausages	Moyano et al., 2011
8	<i>Lactobacillus lactis</i>	Buttermilk and cheese	Madigan M and Martinko J, 2005
9	<i>Lactobacillus brevis</i>	Sauerkraut and pickles	Pavlova et al., 2012
10	<i>Lactobacillus casei</i>	Dairy production and Yakult	Banks et al., 2004
11	<i>Lactobacillus farciminis</i>	Production of nitric oxide	Jean Fioramonti and Roquettes, 2007
12	<i>Lactobacillus paracasei</i>	Dairy product fermentation and probiotics.	Orlando et al., 2012
13	<i>Lactobacillus crispatus</i>	Hydrogen peroxide (H ₂ O ₂) producing beneficial micro biota	Antonio and Hillier, 2003
14	<i>Bifidobacterium bifidum</i>	Mother milk	https://en.wikipedia.org/wiki/Bifidobacterium_bifidum
15	<i>Bifidobacterium infantis</i>	Produces an acid that kills harmful organisms	Edward, 2015
16	<i>Bifidobacterium adolescentis</i>	Synthesizes various B vitamins	Arunachalam, 1999; Pompei et al., 2007
17	<i>Bifidobacterium longum</i>	Fresh milk tofu and pineapple juice	Phoem,2015
18	<i>Bifidobacterium breve</i>	Dietary supplements - capsules, powder, tablets	www.danisco.com/dietarysupplements
19	<i>Bifidobacterium lactis</i>	Inhibit the toxic effects of gliadin	Lindfors et al., 2008
20	<i>Bifidobacterium animalis</i>	Fermentation of Baltic Herring	Pavels Semjonovs et al., 2015
21	<i>Streptococcus salivarius</i>	Chemical-ionization, Synthesis of a broth levan	Peter J. Simms, 1990
22	<i>Streptococcus thermophilus</i>	Cheese manufacture	Daire Stokes et al., 2001
23	<i>Propionibacterium</i>	Acne treatment.	Ewa Jo nczyk-Matysiak et al., 2017
24	<i>Leuconostoc mesenteroides</i>	Production de-glucose-1- phosphate and various sugar	Dols et al., 1997
25	<i>Bacillus clausii</i>	Detergent	Han-Seung Joo and Chung-Soon Chang, 2006
26	<i>Bacillus subtilis</i>	Plant growth and yield, Formation of antibiotic	Kilian et al., 2000.
27	<i>Bacillus licheniformis</i>	Starch liquefaction,	Jan Pen et al.,1992,
28	<i>Bacillus cereus</i>	Pasteurised milk, Chilled dairy products, Alkaline protease, Rice	Zwietering et al., 1996; Yamada et al.,1999
29	<i>Enterococcus mundtii</i>	Biopreservatives in Cheese and meat products, infant milk formulations as substrate	Zendo et.al.,2005; Botes et. al., 2008
30	<i>Enterococcus faecium</i>	Fermented milk product	Agerbaek et al.,1995
31	<i>Saccharomyces cerevisiae</i>	Folate biofortification	Daimmo et al., 2012
32	<i>Saccharomyces boulardii</i>	Lyophilized form of freeze-dried form, Dosage- 250 mg for twice a day for over 2 months child.	Marcia et al., 2009
33	<i>Candida pintolopesii</i> , <i>C. saitoana</i> and <i>S. cerevisiae</i>	Animal feeds as probiotic additives	Bovill et al., 2001; Leuschner et al., 2004

2. Practical Application

Use of Paraprobiotic is an advanced expected technique to overcome the human immune system problems. It triggers a more beneficial biological reaction in human system, which

contributes to a healthy life by the proper production of metabolic cells. Thus, food technologists are working towards the implementation of paraprobiotics in the global marketplace by espousing various methods to improve the food habitats efficiencies. Paraprobiotics has potential application in food

science and nutritional division such as dietary supplements, dairy products, etc. This review concludes that the probiotics and paraprobiotics products are able to improve the human health in better condition and it may be considered as a bio-food for imminent deficiency issues. Paraprobiotics products can also cure Immunomodulation and Inhibition of pathogens.

3. Mechanism of probiotics

Probiotic mechanisms which includes improvement of the epithelial wall, better linkage to intestinal mucosa, and hindering the pathogen adhesion, prohibiting of pathogenic microorganisms, creation of anti-microbial substances and modification in functional activity of the immune system. Probiotics microorganism used to hold intestinal mucus and epithelial cells has considered as one of the most essential assortment principles in bio food. Linkage of intestinal mucosa may avoid the removal of probiotic cells and allowing temporary colonization, immune modulation, and competitive pathogens rejection. Various different techniques used in the bond analyses of probiotics. The more cooperative tests are assessing the linkage to epithelial cell and intestinal mucus. Endogenous bacteria attached to epithelial cells and intestinal mucosa (Yuan Kun Lee and Seppo Salminen, 2009).

4. What is Paraprobiotics?

As per FAO/WHO, probiotics use live microorganism, viable, culturable, and immunologically active microorganism which gives health benefits to the human beings (Adams, 2010; Ananta and Knorr, 2009). The term 'Paraprobiotics' defined as the use of inactivated microorganism or cell fraction to confer a health benefit to the consumer (Taverniti and Guglielmi, 2011) also referred as "*inactivated probiotics*" and "*ghost probiotics*" (Tsilingiri and Rescigno, 2013; Tsilingiri et al., 2012). The term "*postbiotics*" also proposed defined as "non-viable microbial products or metabolic byproduct from probiotic microorganism that cause biological activity in host". Therefore, "paraprobiotics" are also called as "postbiotics" (Patel and Denning, 2013). Production of paraprobiotics required inactivated microorganism, for that several methods are used. In those methods, microorganism becomes partially non-viable due to its physiological changes in microorganism. Sometime, it becomes completely non-viable because of different cell structures like DNA break, cell membrane and cell envelope damages in microorganism. This has been determined by the viability of microorganism using fluorescent dyes which show the viability, characteristics, and differentiates biological parameter like intracellular activity, cytoplasmic enzymes and integrity of the membrane. The main dye used in determination of cell viability like Carboxy fluorescein diacetate (cFDA), Fluorescein diacetate (FDA), Hydroethidine, propidium iodide (PI) and SYTO-9 (Ananta and Knorr, 2009; Doherty et al., 2010).

5. Inactivation method of microorganism

Paraprobiotics contain bacteria concentrations between 1×10^5 to 1×10^{14} per doses (Raz and Rachmilewitz, 2005). Inactivation of probiotic microorganism can be carried out by various methods are used to produce paraprobiotics product. Commercial methods like Thermal treatment (TT), (Chiu et al., 2013) High pressure processing (HPP), (Ananta and Knorr,

2009) Ultraviolet rays (UV), (Lopez, et al., 2008; Van Hoffen et al., 2010) Irradiation (I), (Kamiya et al., 2006) Sonication (S), (Shin et al., 2010) and other methods like Pulsed Electric Field (PEF), Ohmic heating, supercritical CO₂, Drying, and change in pH. This method can cause destruction of the microorganism.

6. Methodology

6.1. Thermal Treatment

Most common methods for inactivation of microorganism for the production of paraprobiotics is thermal treatment. In this method, heat is applied on microorganism for a specific period. This time period varies from organism to organism according to their stability and characteristics like the type of microorganism, vegetative cell or spores, growth medium, growth stage, prior exposition, stress, pH, water activity, mode of heating etc., (Juneja and Sofos, 2002). This treatment is based on thermal resistance of microorganism, which is given by two parameters D and Z value. The D value represents the time required to reduce 90% of microbial population at a specific temperature and the Z value represents the temperature range for a reduction in the D value (Awuah et al., 2007).

In paraprobiotics, thermal treatment results in an increased the cell smoothness or roughness which result in immune modulating properties. If, high temperature is applied it results in more roughness. Researchers said that heat treatment improve or reduce the effect of paraprobiotics. The authors reported that decrease in body weight and viral load in the lungs of mice has infected by the flu virus. This has been cured by supplementing the paraprobiotics organism, i.e., *L. gasseri* (treated at 90° C for 5 min) is used for inactivation of flu virus. Other animals are treated by same paraprobiotic organism treated at 70° C for 30 min and suspended in saline solution (NaCl). More diversity in heat inactivation condition of probiotics results in production of paraprobiotics. The time and temperature vary from 5 to 60 min at 60 to 121° C at specific condition depends on the microorganism used in paraprobiotics (Kawase et al., 2012; Ou et al., 2011).

6.2. Ionizing radiation

Application of electromagnetic waves or electron is known as ionizing radiation on probiotics for inactivation to produce paraprobiotics. The radiation sources are gamma rays emitted from radioisotopes Co60 and Cs137, high energy electron and X-rays. The amount of radiation absorbed is measured in Grays (Gy) (Farkas and Mohacsi-Farkas, 2011; Ibrahim, 2013). The 2-10 kGy refer to ionizing radiation (Farkas, 1998). Treatment up to 10 kGy is considered as safe ionizing radiation (Farkas, 2006; Mansur, et al., 2014). In this method, microorganism inactivation takes place by damaging the nucleic acids directly or indirectly. Absorption of radiation energy per target molecule takes place in direct damage, and in indirect damage interaction between reactive species formed by radiolysis of water like hydroxyl radical (Manas and Pagan, 2005). But, now a day production of paraprobiotics from ionizing radiation is banned. Because of the few drawbacks in ionizing radiation, they are over usage/over exposure of organisms in ionizing results in production of toxic biological products due to change in DNA or RNA structure. The 1,2 M rad using a cesium 137 source at 8 Gy/min for overnight is a

more effective treatment of Colitis in animal (Raz and Rachmiewitz, 2005). Cobalt 60 source for 20 h at 8.05 Gy/min used to inactivate *L. reuteri* (5×10^9 bacteria/ml in MRS broth).

6.3. Ultraviolet rays

The UV rays are categorized under non-ionizing radiation and it has germicidal property (Lado and Yousef, 2002). Electromagnetic spectrum ranges from 200 to 400 nm are considered under UV rays. It also contains short ranges from 200 to 280 nm, medium from 280 to 320 nm and long UV waves from 320 to 400 nm. Mostly, short wave is used for inactivation of microorganism. Some of the major advantages which encountered in UV methods are applied to disinfect the air, surface, water, etc., It also inactivates the vegetative bacterial cell and spores (Gayan et al., 2013). It has minimum loss of sensory qualities without toxic effect takes place. Low energy is required as compared to other methods (Gayan et al., 2014). This method used for inactivation of microorganism by protein denaturation and production of DNA photoproduct, i.e., Pyrimidine dimer formed between adjacent pyrimidine on the same DNA strand and interrupt DNA transcription or translation may cause mutagenesis and death of cell occur (Franz et al., 2009). The production of paraprobiotics required to expose 5 to 30 min in UV.

6.4. High Hydrostatic Pressure

In this method, high pressure is applied at varying times on microorganism for inactivation. This is the anticipated new technology in the food industries for inactivation of microorganism. The HHP is employed at different ranges of pressure, i.e., 100 to 1200 MPa, but direct pressure is not applied on microorganism due to the fluid medium is getting transferred or spill out from the reactor. By direct pressure method may cause damages or demises the microorganism (Chawla, et al., 2011; Rendueles et al., 2011). Microorganism are inactivated by the homogenization process in that high pressure is applied which varies from 30 to 350 MPa. Inactivation microorganism happened by increasing the pressure and shear stress leads to rupture of microbial cell (Diels and Michiels, 2016). The pressure ranges from 20 to 180 MPa is used to reduce the microbial growth and suppress protein synthesis. At 180 MPa pressure, cell lose their viability and above 300 MPa irreversible protein denaturation takes place and inactivate the vegetative cell and bacteriophages (Lado and Yousef, 2002). Enzymes required high pressure to inactivate. Comparison between thermal inactivation and high-pressure inactivation is 60 to 75°C for 15-300s and 400 to 600 MPa at 37 °C for 10 min respectively for cells of *L. rhamnosus* in phosphate buffer solution (PBS) at concentration of 10⁹ CFU/ml. If above 400 MPa high pressure leads to lethal effect on cell (Ananta and Knorr, 2009).

6.5. Sonication

Ultrasound is used to inactivate microorganism. It is a physical method to break the intermolecular attraction. Ultrasound frequency is more than an outpost of human audition (more than 16 kHz) (Gibson et al., 2008). In this method, gas bubbles are produced in liquid media after explosion of bubbles the temperature and force is created per unit area. Inactivation depends on the nature of ultrasound wave, time, and type of microorganisms used (Butz and

Tauscher, 2002). Physical and chemical change takes place because of intracellular cavity. Inactivation of microorganism takes place due to breaking of cell membrane, cell wall, DNA damage and free radical production (Birmpa, et al., 2003). In sonication, *Bifidobacterium longum* is inactivated by placing in saline solution at 10⁸-10⁹ CFU/ml for 5 min and reduces the cholesterol level (Shin et al., 2010).

6.6. Other methods for inactivation

- 6.6.1. Pulse electric field- High or low voltage is applied to the microorganism placed in between two electrodes for inactivation (Gongora et al., 2004). The electric field in between is 2 to 87 kV/cm. It is used for inactivation of enzymes, spores, yeast, gram positive vegetative cell and gram-negative bacteria (Devlieghere et al., 2004; Gongora-Nieto et al., 2002).
- 6.6.2. Ohmic heating- It is also known as joule heating. Heating is occurring by passing the electric current in it. Organisms have well electrical resistance property due to that it capable to produce heat within it. In this process, it reduces the entry of energy and thermal damage (Pereira and Vicente, 2010).
- 6.6.3. Supercritical CO₂ Technology- High concentration CO₂ is used at critical temperatures and pressure. The critical temperature is 31.1°C and pressure is 7.38 MPa. It is more profitable, because of CO₂ is cheap, environmentally safe, non-toxic, and use below 20 MPa pressure (Efaq et al., 2015; Garcia et al., 2007; Yuk and Geveke, 2011).
- 6.6.4. Dehydration- In this process, removal of water from microorganism take place. Therefore, water activity decreases and microorganism growth is stopped. Removal of water by lyophilization, spray drying and osmosis are used. In lyophilization, the microorganism is frozen and placed in a vacuum and spray drying which results in powder form. The drying temperature used for spray dryer is 150-180 °C so, the decline of microorganism occurs.
- 6.6.5. pH-microorganism grows in specific pH it may be above or below the neutral level of microorganism. Therefore, changing the pH will result in inactivation of cells (Foerst et al., 2011; Ananta et al., 2005; Peighambardoust et al., 2011).

7. Application of Paraprobiotics

7.1. Immunomodulation

A chemical agent that modifies the immune response or the functioning of the immune system is known as immunomodulation. Immune system divided into innate and adaptive system. Innate system works with the help of white blood cells. Adaptive system works with the help of lymphocytes B and T. (Bermudez-Brito et al., 2012; Oelschlaeger T.M. 2010). *Paraprobiotics* act on the immune system and alter them. *L. gasseri* TMC0356 and *L. plantarum* L-137 paraprobiotics show vitro immunomodulation effect. *L. plantarum* L-137 help in stimulating the production of IL-12p40 (Miyazawa et al., 2011)

7.2. Inhibition of pathogens

Paraprobiotics protect the host against infection. Protection mechanism is direct inactivation of pathogens by the production of lactic acid, bacteriocin, etc. (Bermudez-Brito et al., 2012) *L. plantarum* b240 protect from the infection of *Salmonella enterica* sarovar, *Typhimurium* (Ishikawa et al., 2010) and *Ln. mesenteroides* 1RM3 protect from irruption and infection by *Listeria monocytogenes* (Nakamura et al., 2012), *L. plantarum* L-137 protected from H1N1 flu virus and *L. plantarum* 062 also (Maeda et al., 2009).

8. Additional use of Paraprobiotics

Some of the other function of paraprobiotics are Recovery of intestinal injuries, Reduction of bacterial translocation and preservation of the intestinal barrier, Treatment of diarrhea, Reducing lactose intolerance, Cholesterol reduction, Respiratory diseases, Improvement in alcohol induced liver diseases, Inhibition of growth of cancer, Treatment of atopic dermatitis, Modulation of response to visceral pain, Treatment of colitis, Suppression of some age associated manifestations, and Inhibition of dental caries. **Table 2** describes the various paraprobiotics Microorganism/Product for that the Concentration, Inactivation methods and Condition of inactivation.

Table-2

Preparation of paraprobiotics Microorganism/Product, Concentration, Inactivation methods, Condition of inactivation, and Reference List

Microorganism/ Product	Concentration	Method of inactivation	Condition of inactivation	Reference
<i>Bifidobacterium breve</i> Yakult (BbrY) and <i>Bifidobacterium bifidum</i> Yakult (BbiY)	--	Heat	100 ° C for 30 min	Kamilya et al., 2015
<i>Bifidobacterium longum</i> SPM1207	10 ⁸ -10 ⁹ CFU/ml	Sonication	Sonication for 5 min	Shin et al., 2010
Capsules containing <i>L. acidophilus</i> LB	5 billions/ capsule	Heat	–	Xiao et al., 2002
<i>E. faecium</i> JWS 833	--	Heat	110 ° C for 15 min	Choi et al., 2012; Sawada et al., 2016
Fermented Milk containing <i>L. gasseri</i> CP2305	1×10 ¹⁰ bacterial cells per container	Heat	95 ° C for 30 min	Sawada et al., 2016.
<i>L. acidophilus</i> (LAP5, LAF1, LAH7)	1×10 ¹⁰ CFU/ml	Heat	100 ° C for 30 min	Lin et al., 2007.
<i>Lactobacillus brevis</i> SBC8803	1.45×10 ⁹ CFU/mg	Heat	121 ° C for 20 min	Segawa et al., 2008
<i>L. brevis</i> SBC8803	–	Heat	121 ° C for 20 min	Ueno et al., 2011.
<i>L. casei</i> CRL 431	10 ⁹ cells	Heat	80 ° C for 30 min	Villena et al., 2009.
<i>L. gasseri</i> OLL2809	5×10 ⁸ cell/mg	Heat	75 ° C for 60 min	Sashihara et al., 2008.
<i>L. plantarum</i> 06CC2	–	Heat	Boil for 1 hr.	Takeda et al., 2011.
<i>L. plantarum</i> b240	–	Heat	121 ° C for 15 min	Ishikawa et al., 2010
<i>L. reuteri</i>	1×10 ⁹	irradiation heat	80 ° C for 20 min., Cobalt 60 source for 20 hr. at 8.05 Gy/min	Kamiya et al., 2006.
<i>L. rhamnosus</i> GG	10 ⁶ , 10 ⁸ , and 10 ¹⁰ CFU/ml; 10 ⁸ CFU/L	Heat	80 ° C for 20 min.	Bloise et al., 2010; Li et al., 2009.
<i>L. lactis</i> G50	0.5 g/Kg (w/w)	Heat	100 ° C for 30 min	Kimoto-Nira et al., 2009
<i>Leuconostoc mesenteroides</i> 1RM3	9 log CFU/ml	Heat	100 ° C for 10 min	Nakamura et al., 2012.

Skimmed milk supplemented with <i>L. rhamnosus</i> HN001	10 ⁹ microorganism/day	Heat	100 ° C for 15 min	Gill and Rutherford, 2001
Supplementation with <i>L. casei</i> GG	10 ¹⁰⁻¹¹ CFU/g	Heat	85-100 ° C for 10 min	Kaila et al., 1995
Tablets containing <i>L. gasseri</i> OLL2809	1×10 ¹⁰ cells/tablet	Heat	75 ° C for 60 min	Gotoh et al., 2009.
Yogurt containing lactic acid bacteria (<i>L. bulgaricus</i> , <i>S. thermophilus</i> , <i>L. acidophilus</i>) (Mix-LAB)	1× 10 ⁷ , 1 ×10 ⁸ and 1× 10 ⁹ cells/ml	Heat	65 ° C for 60 min	Zeng et al., 2015.

9. Conclusion

Evidence show in this review paraprobiotics acts as probiotics but, mechanism of action is different and give health benefit to human being, cannot cause infection and harmful diseases to the body. It also gives the nutrition to the body. It secretes various enzymes and acids, which is involved in metabolic reaction and it also decreases the deficiency of

vitamins and minerals. The production of paraprobiotic product is cheap and easy. In paraprobiotics, modified microorganism is used. The improvement of paraprobiotics as constituents and their application in food and beverages is important. At the time of processing probiotics are not survive but paraprobiotics are survive.

References

- Adams, C.A. (2010). The probiotic paradox: Live and dead cells are biological response modifier. *Nutrition Research Reviews*, 23, 037-46.
- Agerbaek, M., Gerdas. L.U., Richelsen, B. (1995). Hypocholesterolemia effect of a new fermented milk product in healthy middle-aged men. *Eur J Clin Nutr*, 49, 346-52.
- Amara, A.A., and Shilb. A. (2013). Role of probiotics in health improvement, infection control and disease treatment and management. *Saudi Pharmaceutical Journal*, 23, 107-114.
- Ananta, E., Volkert, M., and Knorr, D. (2005). Cellular injuries and storage stability of spray dried *Lactobacillus rhamnosus* GG. *International Dairy Journal*, 15, 399-409.
- Ananta, E., and Knorr D. (2009). Comparison of inactivation pathway of thermal or high-pressure inactivation *Lactobacillus rhamnosus* ATCC 53103 by flow cytometry analysis. *Food Microbiology*, 26, 542-546.
- Anekella, K., and Orsat, V. (2013). Optimization of microencapsulation of probiotics in raspberry juice by spray drying. *LWT- Food Science and Technology*, 50,17-24.
- Anna Pompei, P., Lisa Cordisco, Alberto Amaretti, Simona Zanoni, Diego Matteuzzi, and Maddalena Rossi. (2007). *Folate Production by Bifidobacteria as a Potential Probiotic Property*. *Applied and Environmental Microbiology*, 73(1), 179-185.
- Antonio, M.A., and Hillier, S.L., (2003). DNA Fingerprinting of *Lactobacillus crispatus* Strain CTV-05 by Repetitive Element Sequence-Based PCR Analysis in a Pilot Study of Vaginal Colonization. *J. Clin. Microbiol*, 41 (5), 1881-7.
- Arora T., Broglia E., Pushpa Kumar D., Lodhi T., Taheri S. (2013) An investigation into the strength of the association and agreement levels between subjective and objective sleep duration in adolescents. *PLoS One* 8: e72406.
- Arun K Panda, Savaram S Rama Rao, Manteta VLN Raju, and Sita S Sharma, (2008). Effect of probiotic (*Lactobacillus sporogenes*) feeding on egg production and quality, yolk cholesterol and humoral immune response of White Leghorn layer breeders. *Journal of the Science of Food and Agriculture*, 88 (1), 43-47.
- Arunachalam and Kantha D. (1999). The Role of Bifidobacteria in Nutrition, Medicine, and Technology. *Nutrition Research*, 19(10), 1559-1597.
- Arvind Kumar and Shafat, S. (2010). Probiotics-Role and Importance In Food Of Animal Origin, Beverage and Food World.
- Auwah, G.B., Ramaswamy, H.S., and Economides, A., (2007). Thermal processing and quality: Principles and overview. *Chemical Engineering and Processing: Process Intensification*, 46, 584-602.
- Banks, J M., Williams, A G., and Williams. (2004). The role of the nonstarter lactic acid bacteria in Cheddar cheese ripening. *International Journal of Dairy Technology*, 57 (2-3): 145-152.
- Bermudez-Brito, M., Plaza-Diaz, J., Munoz-Quezada, S., Comez-Llorente, C., and Gil, A. (2012) Probiotic mechanisms of action. *Annals of Nutrition & Metabolism*, 61,160-174.
- Betoret, N., Puente, L., Diaz, M.J., Garcia, M.J., Gras, M.L., Martínez-Monzó, J., and Fito P., (2003). Development of probiotic-enriched dried fruits by vacuum impregnation. *Journal of Food Engineering*, 56, 273-277.
- Birmpa, A., Sfika, V., & Vantarakis, A. (2013). Ultraviolet light and ultrasound as nonthermal treatments for the inactivation of microorganisms in fresh ready-to-eat foods. *International Journal of Food Microbiology*, 167, 96-102.
- Bloise, E., Torricelli, M., Novembri, R., Borges, L.E., Carrarelli, P., and Reis, F.M. (2010). Heat killed *Lactobacillus rhamnosus* GG modulates urocortin and cytokine release in primary trophoblast cells. *Placenta*, 31, 867-872.
- Bordoni, A., Amaretti, A., Leonardi, A., Boschetti, E., Danesi, F., Matteuzzi, D., Roncaglia, L., Raimondi, S., and Rossi, M. (2013). Cholesterol-lowering probiotics: In vitro selection and in vivo testing of Bifidobacteria. *Applied Microbiology and Biotechnology*, 97, 8273-8281.
- Botes M, Van Reenen C.A., and Dicks L.M. (2008). Evaluation of *Enterococcus mundtii* ST4SA and *Lactobacillus plantarum* 423 as probiotics by using a gastro-intestinal model with infant milk formulations as substrate. *Int J Food Microbiol*, 128(2): 362-70.

21. Bovill, R., Bew, J., and Robinson, S. (2001). Comparison of selective media for the recovery and enumeration of probiotic yeasts from animal feed. *Int J Food Microbiol*, 67(1-2):55-61.
22. Butz, P., and Tauscher, B. (2002). Emerging technologies: Chemical aspects. *Food Research International*, 35, 279-284.
23. Caroline N. de Almada, Carine N. Almada, Rafael Martinez, C.R., and Anderson S. Sant'Ana, (2016). Paraprobiotics: Evidences on their ability to modify biological responses, inactivation methods and perspectives on their application in foods. *Trends in Food Science & Technology*, 58:96–114.
24. Chawla, R., Patil, G.R, and Singh, A.K. (2011). High hydrostatic pressure technology in dairy processing: A review. *Journal of Food Science and Technology*, 48, 260-268.
25. Chiang, S., and Pan, T., (2011). Beneficial effects of *Lactobacillus paracasei* sub sp. *paracasei* NTU 101 and its fermented products. *Applied Microbiology and Biotechnology*, 93, 903-916.
26. Chiu, Y.H., Lu, Y.C., Ou, C.C., Lin, S. L., Tsai, C.C., Huang, C.T., and Meei-Yn Lin. (2013). *Lactobacillus plantarum* MYL26 induces endotoxin tolerance phenotype in Caco-2 cells. *BMC Microbiology*, 13, 1-9.
27. Choi, H.J., Shin, M.S., Lee, S.M., and Lee, W.K. (2012). Immunomodulatory properties of *Enterococcus faecium* JWS 833 isolated from duck intestine track and suppression of listeria monocytogenes infection. *Microbiology and Immunology*, 56, 613-620.
28. Daimmo, M.R, Mattarelli, P., Biavati, B., Carlsson, N.G., and Andlid, T. (2012). The potential of Bifidobacteria as a source of natural folate. *J Appl Microbiol*, 112(5), 975-84.
29. Daire Stokes, Paul Ross, R., Gerald Fitzgerald, and Aidan Coffey. (2001). Application of *Streptococcus thermophilus* DPC1842 as an adjunct to counteract bacteriophage disruption in a predominantly lactococcal Cheddar cheese starter: use in bulk starter culture systems, 10th Meeting of the " Club des Bactéries Lactiques", Lait, 81 (1-2), 327 – 334.
30. Devlieghere, F., Vermeiren, L., and Debevere, J. (2004). New preservation technologies: Possibilities and limitations. *International Dairy Journal*, 14, 273-285.
31. Diels, A.M., and Michiels, C.W., (2016). High pressure homogenization as a non-thermal technique for the inactivation of microorganism. *Critical Reviews in Microbiology*, 32, 201-216, 2016.
32. Doherty, S.B., Wang, L., Ross, R.P., Stanton, C., Fitzgerald, G.F., and Brodtkorb, A. (2010). Use of viability staining in combination with flow cytometry for rapid viability assessment of *Lactobacillus rhamnosus* GG in complex protein matrices. *Journal of microbiological Methods*, 82, 301-310.
33. Dols, M., Chraïbi, W., Remaud-Simeon, M., Lindley, N D., and Monsan, P F. (1997). Growth and energetics of *Leuconostoc mesenteroides* NRRL B-1299 during metabolism of various sugars and their consequences for dextranucrase production. *Appl Environ Microbiol*. 63(6), 2159–2165.
34. Ducrotre, P., Sawant, P., and Jayanthi, V. (2012). Clinical trial: *Lactobacillus plantarum* 299v (DSM9843) improve symptoms of irritable bowel syndrome. *World journal of Gastroenterology*, 18, 4012-4016.
35. Edward Group DC, NP, DACBN, DCBCN, DABFM. (2015). Bifidobacterium infants: A Healthy Probiotic Strain. <http://www.globalhealingcenter.com/natural-health/bifidobacterium-infantis-the-health-benefits-of-probiotics>.
36. Efaq, A.N., Rahman, N. N. A., Nagao, H., Al-Gheethi, A.A., Shahadat, M., and Kadir, M. O. A. (2015). Supercritical carbon dioxide as non-thermal alternative technology for safe handling of clinical wastes. *Environmental Processes*, 2, 797-822.
37. Ewa Jończyk-Matysiak, Beata Weber-D abrowska, Maciej Zaczek, Ryszard Mi edzybrodzki, Sławomir Letkiewicz, Marzanna Łusiak-Szelchowska and Andrzej Górski. (2017). Prospects of Phage Application in the Treatment of Acne Caused by *Propionibacterium acnes*. *Front Microbiol*, 8:164.
38. Farkas, I. (1998). Irradiation as a method for decontaminating food. A review *International Journal of Food Microbiology*, 44, 189-204.
39. Farkas, J. (2006). Irradiation for better foods. *Trends in Food Science & Technology*, 17, 148-152.
40. Farkas, J., and Mohacsi-Farkas, C. (2011). History and future of food irradiation. *Trends in Food Science & Technology*, 22, 121-126.
41. Foerst, P., and Kulozik, U. (2011). Modelling the dynamic inactivation of the probiotic bacterium *L. Paracasei* ssp. *Paracasei* during a low-temperature drying process based on stationary data in concentrated systems. *Food and Bioprocess Technology*, 5, 2419-2427.
42. Franz, C., Specht, I., Cho, G. S., Graef, V., and Stal, M. (2009). UV-C inactivation equipment based on Dean vortex technology. *Food Control*, 20, 1103-1107.
43. Garcia-Gonzalez, L., Geeraerd, A.H., Spilimbergo, S., Elst, K., Van Ginneken, L., Debevere J. et al., (2007). High pressure carbon dioxide inactivation of microorganism in food: The past, The present and the future. *International Journal of Food Microbiology*, 117, 1-28.
44. Gayan, E., Alvarez, I., and Condon, S. (2013). Inactivation of bacterial spores by UV-C light. *Inactivation Food Science & Emerging Technology*, 19, 140-145.
45. Gayan, E., Condon, S., and Alvarez, I., (2014). Biological aspects in food preservation by ultraviolet light. A review *Food and Bioprocess Technology*, 7, 1-20.
46. Gibson, J.H., Hai, D., Yong, N., Farnood, R.R., and Seto, P. (2008). A literature review of ultrasound technology and its application in wastewater disinfection, 43, 23-35.
47. Gill, H.S., and Rutherford, K.J. (2001). Viability and dose response studies on the effect of the immune enhancing lactic acid bacterium *Lactobacillus rhamnosus* in mice. *British Journal of Nutrition*, 86,285-289.
48. Gongora-Nieto, M. M., Sepúlveda, P. P., Pedrow, P., Barbosa-C anovas, G. V., & Swanson, B. G. (2002). Food processing by pulsed electric Fields: Treatment delivery, inactivation level, and regulatory aspects. *LWT - Food Science and Technology*, 35, 375-388.
49. Gongora-Nieto, M.M., Pedrow, P.D., Swanson, B.G., and Barbosa-Canovas, G.V. (2004). Use of circuit analysis simulation in pulsed electric fields food processing. *Journal of Food Engineering*, 61, 413-420.
50. Gotoh, M., Sashihara, T., Ikegami, S., Yamaji, T., Kino, K., Orii, N., Taketomo, N., Okubo, K. (2009). Efficacy of oral administration of a heat killed *Lactobacillus gasseri* OLL2809 on patients of Japanese cedar pollinosis with high Japanese cedar pollen specific IgE. *Bioscience. Biotechnology and Biochemistry*, 73, 1971-1977.
51. Han-Seung Joo and Chung-Soon Chang. (2006). Production of an oxidant and SDS-stable alkaline protease from an alkalophilic *Bacillus clausii* I-52 by submerged fermentation: Feasibility as a laundry detergent additive. *Enzyme and Microbial Technology*, 38 (1–2), 176–183.
52. Hsieh, F., Lee, C., Chen, W., Lu, Y., and Wu, C. (2013). Oral administration of *Lactobacillus reuteri* GMNL-263 improves insulin resistance and ameliorates hepatic steatosis in high fructose-fed rats, *Nutrition & Metabolism*, 10, 1-14.
53. https://en.wikipedia.org/wiki/Bifidobacterium_bifidum "Bifidobacterium bifidum"
54. Ibrahim, H. M. (2013). Prediction of meat and meat products by gamma rays, electron beams and X-ray irradiations-A

- Review. International Journal of Agricultural Sciences, 3, 521-534
55. Ishikawa, H., Kutsukake, E., Fukui, T., Sato, L., Kurihara, T., Okada, N., Danbara, H., Toba, M., Kohda, N., Maeda, Y., and Matsumoto, T. (2010). Oral administration of heat-killed *Lactobacillus plantarum* strain b240 protected mice against *Salmonella enterica* serovar typhimurium. *Bioscience, Biotechnology, and Biochemistry*, 74, 1338-1342.
 56. Jan Pen, Lucy Molendijk, Wim J. Quax, Peter C. Sijmons, Albert J. J. van Ooyen, Peter J. M. van den Elzen, Krijn Rietveld and André Hoekema. (1992). Production of Active *Bacillus licheniformis* Alpha-Amylase in Tobacco and its Application in Starch Liquefaction. *Biotechnology (N Y)*. 10(3):292-6.
 57. Jean Fioramonti, Roquettes (FR), Lionel Bueno, Aussonne (FR), Vassilia, Theodorou, Portet-sur-Garonne (FR), Florence Lamine, Toulouse (FR). (2007). Use of *Lactobacillus farciminis* for the prevention or treatment of digestive pathologies. Patent.
 58. Juan Manuel Palomino, Julia Toledo del Árbol, Nabil Benomar, Hikmate Abriouel, Magdalena Martínez Cañamero, Antonio Gálvez, and Rubén Pérez Pulido. (2015). Application of *Lactobacillus plantarum* Lb9 as starter culture in caper berry fermentation. *LWT - Food Science and Technology* 60 (2), Part 1, 788–794.
 59. Juneja, V., and Sofos, J.N. (2002). Thermal inactivation of microorganism, In M. Dekker (Ed.) *Control of foodborne microorganism.*, New York: Food Science and Technology, 13-15.
 60. Kaila, M., Isolauri, E., Saxelin, M., Arvilommi, H., and Vesikari, T. (1995). Viable versus inactivation *Lactobacillus* strain GG in acute rotavirus diarrhea. *Archives of Disease in Childhood*, 72, 51-53.
 61. Kamilya, D., Baruah, A., Sangma, T., Chowdhury, S., & Pal, P. (2015). Inactivated probiotic bacteria stimulate cellular immune responses of catla, (Hamilton) in vitro. *Probiotics and Antimicrobial Proteins*, 7, 101-106.
 62. Kamiya, T., Wang, L., Forsythe, P., Goettsche, G., Mao, Y., et al., (2006). Inhibitory effects of *Lactobacillus reuteri* on visceral pain induced by colorectal distension in Sprague-Dawley rats. *Gut*, 55, 191-196.
 63. Kawase, M., He, F., Kubota, A., Yoda, K., Miyazawa, K., and Hiramatsu, M. (2012). Heat killed *Lactobacillus gasseri* TMC0356 protects mice against influenza virus infection by stimulating gut and respiratory immune responses. *FEMS Immunology and Medical Microbiology*, 64, 280-288.
 64. Kilian, M., Steiner, U., Krebs, B., Junge, H., Schmiedeknecht, G., and Hain R. (2000). FZB24® *Bacillus subtilis* – mode of action of a microbial agent enhancing plant vitality. *Pflanzenschutz-Nachrichten Bayer*, 53(1),72-93.
 65. Kimoto-Nira, H., Mizumachi, K., Okamoto, T., Sasaki, K., and Kurisaki, J. (2009). Influence of long-term consumption of a *Lactococcus lactis* strain on the intestinal immunity and intestinal flora of the senescence-accelerated mouse. *The British Journal of Nutrition*, 102, 181-185.
 66. Kollath, W. (1953). Nutrition and the tooth system; general review with special reference to vitamins. *Deutsche zahnärztliche Zeitschrift*, 8(11), Suppl 7.
 67. Krasaekoopt, W., and Watcharapoka, S., (2014). Effect of addition of insulin and galacto oligosaccharide on the survival of microencapsulated probiotics in alginate beads coated with chitosan in simulated digestive system, Yogurt and fruit juice. *LWT-Food Science and Technology*, 57, 761-766.
 68. Lado, B.H., and Yousef, A.E. (2002). Alternative food-preservation technologies: Efficacy and mechanisms. *Microbes and Infection*, 4, 433-440.
 69. Leuschner, J., Bew, P., Fourcassier, G., and Bertin. (2004). Validation of the official control method based on polymerase chain reaction (PCR) for identification of authorized probiotic yeast in animal feed. *Syst. Appl. Microbiol.* 27, 492–500.
 70. Li, J., Zhang, W., Wang, C., Yu, Q., Dai, R., and Pei, X. (2012). *Lactococcus lactis* expressing food-grade β -galactosidase alleviates lactose intolerance symptoms in post-weaning Balb/c mice., *Applied Microbiology and Biotechnology*, 96, 1499-1506.
 71. Li, N., Russell, W.M., Douglas-Escobar, M., Hauser, N., Lopez, M., and Neu, J. (2009). Live and heat killed *Lactobacillus rhamnosus* GG: Effect on proinflammatory and anti-inflammatory cytokines/chemokines in gastrostomy-fed infants rats. *Pediatric Research*, 66, 203-207.
 72. Lilly DM and Stillwell Rh. (1965). Probiotics: growth promoting factors produced by microorganisms. *Science*, 147(3659),747-8.
 73. Lin, W.-H., Yu, B., Lin, C.-K., Hwang, W.-Z., & Tsen, H.-Y. (2007). Immune effect of heat-killed multi strain of *Lactobacillus acidophilus* against *Salmonella typhimurium* invasion to mice. *Journal of Applied Microbiology*, 102, 22-31.
 74. Lindfors, K., Blomqvist, T., Juuti-Uusitalo, K., Stenman, S., Venäläinen, J., Mäki, M., and Kaukinen, K. (2008). Live probiotic *Bifidobacterium lactis* bacteria inhibit the toxic effects induced by wheat gliadin in epithelial cell culture. *Clin Exp Immunol*, 152(3):552-8.
 75. Lopez, M., Li, N., Kataria, J., Russell, M., and Neu, J. (2008). Live and ultraviolet inactivated *Lactobacillus rhamnosus* GG decrease flagellin-induced Interleukin-8 production in Caco-2 cells. *The Journal of Nutrition*, 2264-2268.
 76. Madigan, M., and Martinko, J. (2005). *Brock Biology of Microorganisms* (11th ed.). Prentice Hall.
 77. Maeda, N., Nakamura, R., Hirose, Y., Murosaki, S., Yamamoto, Y., Kase, T., et al., (2009). Oral administration of heat-killed *Lactobacillus plantarum* L-137 enhances protection against influenza virus infection by stimulation of type 1 interferon production in mice. *International Immunopharmacology*, 9, 1122-1125.
 78. Manas, P., and Pagan, R. (2005). Microbial inactivation by new technologies of food preservation. *Journal of Applied Microbiology*, 98, 1387- 1399.
 79. Mansur, A.R., Yu, C.C., and Oh, D. (2014). Efficiency of gamma irradiation to inactivate growth and fumonisin production of *Fusarium moniliform* on corn grain., *Journal of Microbiology and Biotechnology*, 24, 209-216.
 80. Marcia L. Buck, and Pharm, D. (2009). *Saccharomyces Boulardii* as a Probiotic for Children, *FCCP Pediatric Pharm* 15(7).
 81. Miyazawa, K., He, F., Kawase, M., Kubota, A., Yoda, K., and Hiramatsu, M. (2011). Enhancement of immunoregulatory effect of *Lactobacillus gasseri* TMC0356 by heat treatment and culture media., *Letters in Applied Microbiology*, 53, 210-216.
 82. Mohamadi Sani, Ehsani, M.R., Mazaheri Assadi, M. (2005). Application of Reuterin Produced by *Lactobacillus reuteri* DSM 20016 to Inhibit Some Food-borne Pathogens in UF-Feta-Cheese. *Iranian Research Organization for Sci. & Tech. Tehran-Iran*.
 83. Nakamura, S., Kuda, T., An, C., Kanno, T., Takahashi, H., and Kimura, B. (2012). Inhibitory effects of *Leuconostoc mesenteroides* 1RM3 isolated from narezushi, a fermented fish with rice, on *Listeria monocytogenes* infection to Caco-2 cells and A/J mice., *Anaerobe*, 18, 19-24.
 84. Oelschlaeger, T. A. (2010). Mechanisms of probiotic actions: A review. *International Journal of Medical Microbiology*, 300, 57-62.
 85. Orlando, A, Refolo, M. G., Messa, C., Amati, L., Lavermicocca, P., Guerra, V., and Russo, F. (2012). Antiproliferative and Proapoptotic Effects of Viable or Heat-

- Killed IMPC2.1 and GG in HGC-27 Gastric and DLD-1 Colon Cell Lines, Nutrition and Cancer. 64 (7).
86. Ou, C.C., Lin, S.L., Tsai, J.J., and Lin, M.Y. (2011). Heat Killed lactic acid bacteria enhance immunomodulatory potential by skewing the immune response toward the polarization., Journal of Food Science, 76, M260-M267.
 87. Patel, R.M., and Denning, P.W. (2013). Therapeutic use of prebiotics, probiotics, and postbiotics to prevent necrotizing enterocolitis: What is the current evidence? Clinics in Perinatology, 40, 11-25.
 88. Pavels Semjonovs, Lilija Auzina, Dagnija Upite, Mara Grube, Karlis Shvirksts, Raimonds Linde, Ilze Denina, Artis Bormanis, Andris Uptis, Maija Ruklisha, Elga Biruta Parele, Juris Gailitis, Laima Silina, Emils Kozlinskis, Mara Marauska, Aleksejs Danilovich and Roberts Dlohi. (2015). Application of *Bifidobacterium animalis* subsp. lactis as Starter Culture for Fermentation of Baltic Herring (*Clupea harengus membras*) Mince.
 89. Pavlova, S. I., Kilic, A. O., Kilic, S. S., So, J. S., Nader Macias, M. E., Simoes, J. A., and Tao, L. (2002). Genetic diversity of vaginal *lactobacilli* from women in different countries based on 16S rRNA gene sequences. Journal of applied microbiology, 92(3), 451-459.
 90. Peighambardoust, S. H., Golshan Tafti, a., & Hesari, J. (2011). Application of spray drying for preservation of lactic acid starter cultures: A review. Trends in Food Science & Technology, 22, 215-224.
 91. Pereira, R.N., and Vicente, A.A. (2010). Environmental impact of novel thermal and nonthermal technologies in food processing. Food Research International, 43, 1936-1943.
 92. Peter J. Simms, Walter J. Boyko, John R. Edwards. (1990). The structural analysis of a levan produced by *Streptococcus salivarius* SS2. Carbohydrate Research, 208, 193-198.
 93. Philippe Ducrotté, Prabha Sawant, and Venkataraman Jayanthi. (2012). Clinical trial: *Lactobacillus plantarum* 299v (DSM 9843) improves symptoms of irritable bowel syndrome. World J Gastroenterol 18(30), 4012–4018.
 94. Phoem, AN. (2015). Applications of Microencapsulated Bifidobacterium Longum with Eleutherine Americana in Fresh Milk Tofu and Pineapple Juice. Nutrients, 7(4):2469-84.
 95. Pompei, Anna, et al (2007) Folate Production by Bifidobacteria as a Potential Probiotic Property. Applied and Environmental Microbiology, 73(1), 179-185.
 96. Raz, E., and Rachmilewitz, D. (2005). Inactivated probiotic and methods of use thereof. Patent.
 97. Rendueles, E., Omer, M.K., Alvseike, O., Alonso-Calleja, C., Capita, R., and Prieto, M. (2011). Microbiological food safety assessment of high hydrostatic pressure processing: A review. LWT- Food Science and Technology, 44, 1251-1260.
 98. Rijkers, GT., Bengmark, S., Enck, P., Haller, D., Herz, U., Kalliomäki, M., Kudo, S., Lenoir-Wijnkoop, I., Mercenier, A., Myllyluoma, E., Rabot, S., Rafter, J., Szajewska, H., Watzl, B., Wells, J., Wolvers, D., Antoine, JM. (2010). Guidance for substantiating the evidence for beneficial effects of probiotics: current status and recommendations for future research. J Nutr, 140(3), 671S–676S.
 99. Rokka, S., and Rantamäki, P. (2010). Protecting probiotic bacteria by microencapsulation: Challenges for industrial application. European Food Research and Technology, 231, 1-12.
 100. Rossana Liguori, Carlos Ricardo Soccol, Luciana Porto de Souza Vandenbergh, Adenise Lorenci Woiciechowski, Elena Ionata, Loredana Marcolongo, and Vincenza Faraco, (2015). Selection of the Strain *Lactobacillus acidophilus* ATCC 43121 and Its Application to Brewers, Spent Grain Conversion into Lactic Acid, BioMed Research International.
 101. Santiago Ruiz-Moyano, Alberto Martín, María José Benito, Alejandro Hernández, Rocio Casquete, María de Guía Córdoba. (2011). Application of *Lactobacillus fermentum* HL57 and *Pediococcus acidilactici* SP979 as potential probiotics in the manufacture of traditional Iberian dry-fermented sausages. Food Microbiology 28 (5), 839–847.
 102. Sashihara, T., Ikegami, S., Sueki, N., Yamaji, T., Kino, K., Taketomo, N., et al., (2008). Oral administration of heat-killed *Lactobacillus gasseri* OLL2809 reduces cedar pollen antigen-induced peritoneal eosinophilia in Mice. Allergology International, 57, 397-403.
 103. Sawada, D., Sugawara, T., Ishida, Y., Aihara, K., Aoki, Y., and Takehara, I. (2016). Effect of continuous ingestion of a beverage prepared with *Lactobacillus gasseri* CP2305 inactivated by heat treatment on the regulation of intestinal function. Food Research International, 79, 33-39.
 104. Segawa, S., Wakita, Y., Hirata, H., and Watari, J. (2008). Oral administration of heat killed *Lactobacillus brevis* SBC8803 ameliorates alcoholic liver disease in ethanol-containing diet-fed C57BL/6N mice. International Journal of Food Microbiology, 128, 371-377.
 105. Shin, H.S., Park, S.Y., Lee, D.K., Kim, J.R., et al., (2010). Hypocholesterolemic effect of sonication-killed *Bifidobacterium longum* isolated from healthy adult Koreans in high cholesterol fed rats. Archives of Pharmacal Research, 33, 1425-1431.
 106. Shoichi Yamada, Eiji Ohashi, Norio Agata, And Kasthuri Venkateswaran, (1999). Cloning and Nucleotide Sequence Analysis of *gyrB* of *Bacillus cereus*, *B. thuringiensis*, *B. mycoides*, and *B. anthracis* and their Application to the Detection of *B. cereus* in Rice. Applied and Environmental Microbiology, 1483–1490.
 107. Stavros Plessas, Ann Fisher, Katerina Koureta, Costas Psarianos, Poonam Nigam, Athanasios A. Koutinas. (2008). Application of *Kluyveromyces marxianus*, *Lactobacillus delbrueckii ssp. bulgaricus* and *L. helveticus* for sourdough bread making. Food Chemistry, 106 (3),1:985–990.
 108. Suraji A. Senanayake, Sirimali Fernando, Arthur Bamunuarachchi & Mariam Arsekularatne. (2013). Application of *Lactobacillus acidophilus* (LA 5) strain in fruit-based ice cream. Food Science & Nutrition, 1(6): 428– 431.
 109. Takeda, S., Takeshita, M., Kikuchi, Y., Dashnyam, B., Kawahara, S., Yoshida, H., et al., (2011). Efficacy of oral administration of heat-killed probiotics from Mongolian dairy products against influenza infection in mice: Alleviation of influenza infection by its immunomodulatory activity through intestinal immunity. International Immuno Morphological, 11, 1976-1983.
 110. Taverniti, V., and Guglielmetti, S., (2011). The immunomodulatory properties of probiotic microorganisms beyond their viability (ghost probiotics: Proposal of paraprobiotic concept). Genes & Nutrition, 6, 261-274.
 111. Tsilingiri, K., and Rescigno, M., (2013). Postbiotics: What else? Beneficial Microbes, 4,101-107.
 112. Tsilingiri, K., Barbosa, T., Penna, G., Capriolo, F., Sonzogni, A., Viale, G., et al. (2012). Probiotic and postbiotic activity in health and disease: Comparison on a novel polarized ex-vivo organ culture model. Gut, 61, 1007-1015
 113. Ueno, N., Fujiya, M., Segawa, S., Nata, T., Tanabe, H., et al., (2011). Heat-killed body of *Lactobacillus brevis* SBC8803 ameliorates intestinal injury in a murine model of colitis by enhancing the intestinal barrier function. Inflammatory Bowel Diseases, 17, 2235-2250.
 114. Van Hoffen, E., Korthagen, N.M., Kivit, S., Schouten, B., Bardeel, B., Duivelshof, A., et al., (2010). Exposure of intestinal epithelial cells to UV-killed *Lactobacillus* GG but not *Bifidobacterium breve* enhance the effector immune response in vitro. International Archives of Allergy and Immunology, 152, 159-168.
 115. Villena, J., Barbieri, N., Salva, S., Herrera, M., and Alvarez, S. (2009). Enhanced immune response to pneumococcal

- infection in malnourished mice nasally treated with heat-killed *Lactobacillus casei*. *Microbiology and Immunology*, 53, 636-646.
116. www.danisco.com/dietarysupplements "Bifidobacterium breve"
117. Xiao, S.D., Zhang, D.Z., Lu, H., Jiang, S.H., Liu, H.Y., Wang, G.S., et al., (2002). Multicenter, randomized, controlled trial of heat-killed *Lactobacillus acidophilus* LB in patient with chronic diarrhea. *Chinese Journal of Digestive Diseases*, 3, 167-171.
118. Yamada, S., Ohashi, E., Agata, N., Venkateswaran, K. (1999). Cloning and nucleotide sequence analysis of *gyrB* of *Bacillus cereus*, *B. thuringiensis*, *B. mycoides*, and *B. anthracis* and their application to the detection of *B. cereus* in rice. *Appl Environ Microbiol* 65(4), 483-90.
119. Yuan Kun Lee and Seppo Salminen. (2009). *Handbook of Probiotics and Prebiotics*.
120. Yuk, H.G., and Geveke, D.J. (2011). Nonthermal inactivation and sublethal injury of *Lactobacillus plantarum* in apple cider by a pilot plant scale continuous supercritical carbon dioxide system. *Food Microbiology*, 28, 377-383.
121. Zendo, T., Eunggruttanagorn, N., Fujioka, S., Tashiro, Y., Nomura, K., Sera, Y., Kobayashi, G., Nakayama, J., Ishizaki, A., and Sonomoto, K. (2005). Identification and production of a bacteriocin from *Enterococcus mundtii* QU 2 isolated from soybean.
122. Zeng, J., Jiang, J., Zhu, W., and Chu, Y., (2015). Heat-killed yogurt-containing lactic acid bacteria prevent cytokine-induced barrier disruption in human intestinal caco-2 cells. *Annals of Microbiology*, 66, 171-178.
123. Zwietering, M.H., De Wit, J.C., Notermans, S. (1996). Application of predictive microbiology to estimate the number of *Bacillus cereus* in pasteurized milk at the point of consumption. *International Journal of Food Microbiology*, 30, 55-70.