

Impact of Iron Stress on Oxidative Metabolism in Green Gram Plants

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ABSTRACT

The effect of lacking and poisonous convergences of iron (Fe) on the defensive job of antioxidative proteins just as cancer prevention agent mixes in green gram (*Vigna radiata* (L) Wilczek) plants. Plants were developed in Hoagland supplement arrangement and treated with four Fe convergences of Fe (10,100,200 and 400 μ M Fe) as FeEDTA. Plants demonstrated most extreme development at 100 μ M Fe supply (control). Iron inadequacy was seen at 10 μ M Fe supply and brought about chlorosis of leaves and diminished centralization of dynamic Fe and chlorophyll content. Apical chlorosis, negligible putrefaction and decline in chlorophyll fixation was likewise seen in plants treated with 200 and 400 μ M Fe. High lipid peroxidation (TBARS) and H₂O₂ content in leaves were distinguished in Fe inadequate and Fe dangerous plants when contrasted with control.

1. Introduction

Iron is one of the fundamental components for plant development and generation. Iron is a micronutrient and is required by plants in little sums [1]. Fe in the dirt is the fourth bottomless component on earth, yet its sum is low or not accessible for the plants and microorganisms needs, because of low dissolvability of minerals containing Fe in numerous zones of the world, particularly in bone-dry locale with soluble soils. Its solvency and accessibility are entirely variable and it is reliant on soil pH and Eh. The thin productive Fe focus required for cell needs is carefully constrained by natural procedures acting both at the vehicle and the capacity levels. As a change component Fe exists in two oxidative states: ferrous (Fe²⁺) and ferric (Fe³⁺) and it is required in the majority of the cell redox frameworks such as heme proteins including cytochroms, catalase, peroxidase, leg-hemoglobin and iron sulfur proteins including ferredoxin, acotinase and superoxide dismutase. It is along these lines one of the significant metals engaged with the wide scope of organic capacities like electron-transport chains of plant photosynthesis breath and nitrogen digestion. Abundance of Fe²⁺ anyway hurts the plant by repressing the stretching of plant roots and plant development. Albeit most soils are wealthy in Fe, the "iron harmfulness" is related with overflowed soils which are high in ferrous Fe levels, especially in marsh rice generation territories [2] or tea ranches in India[3]. Poisonous quality of Fe can influence distinctive physiological procedures of the plant. It can prompt the improvement of oxidative pressure [4], by expanding the free extreme creation which irreversibly weakens cell structure and harms DNA and proteins [5], [6]. Insurance techniques against Fe poisonous quality inside the plant may include enzymatic instruments that incorporate the activities of cancer prevention agent compounds, for example, superoxide dismutase, catalase, peroxidase and ascorbate peroxidase and glutathione reductase [7], [8]. Non-enzymatic instruments can kill oxygen receptive species through natural mixes, for example, decreased glutathione (GSH), α -tocopherol, ascorbic corrosive (AA) and carotenoids [9]. On one hand Fe in the free or in the approximately bound structure, goes about as a prooxidant

factor, catalyzing the free extreme age through the Fenton response. Then again, a portion of the cell reinforcement catalysts contain iron, either in heme (POD, CAT) or non-heme structure (FeSOD). Remembering the double job of Fe in plant digestion [10] the present examination means to assess the effect of lacking and harmful dimensions of Fe on the oxidative digestion in green gram plants at various development stages.

2. Review of literature

Plants were isolated into roots, stem and leaves, and all out biomass was controlled by broiler drying (70°C) the examples. The tissue Fe fixation decided in roots, stem and completely extended terminal trifoliate leaves was controlled by nuclear assimilation spectrophotometer (Perkin Elmer Analyst 300) in wet corrosive HNO₃: HClO₄ (10 : 1) digests. Chlorophyll (a + b) and Carotenoids were separated with 80% CH₃CO and estimated spectrophotometrically (Perkin Elmer UV/VIS Lambda Bio 20) as portrayed by strategy for [11]. All out dynamic Fe content in leaves was estimated following the technique for [12]. Ascorbate was separated with 10% trichloroacetic corrosive (TCA) and tested by [13] by following the decrease of Fe³⁺ to Fe²⁺ by ascorbic corrosive and estimating the shading force of the Fe²⁺ - α , α bipyridyl complex at 525 nm. Nonprotein thiols were assessed by the technique for [14]. Leaves were at last cleaved and ground with prechilled mortar and pestle in 5% sulfosalicylic corrosive. After centrifugation, the response was done with 10 mM dithiois 2-nitrobenzoic corrosive (DTNB) and 0.1 mM glutathione diminished (GSH). After 15 min, the shading force of the concentrate was estimated with spectrophotometer at 412 nm. Lipid peroxides were estimated as far as thiobarbituric corrosive receptive substances (TBARS) as depicted by [15]. Crisp leaf material was separated with 1% trichloroacetic corrosive. The supernatant was centrifuged at 10000 g for 10 min and treated with 0.5% thiobarbituric corrosive broke up in TCA. The response blend was brooded in bubbling water shower for 30 min and TBARS were estimated spectrophotometrically at 532 nm in the wake of changing for non-explicit absorbance at 600 nm. Hydrogen peroxide was evaluated by the strategy for [16]. Naturally slashed leaf tissues

were ground in chilled mortar pestle with 100% chilled CH₃CO and centrifuged at 10000 g for 5 min. The supernatant was blended with water and shaken completely. To the supernatant titanium chloride was included trailed by chilled smelling salts. The accelerate was solubilized with the assistance of dainty glass bar and centrifuged. The buildup was washed more than once with CH₃CO to evacuate chlorophyll. The lackluster buildup was broken up in 2N H₂SO₄. The shading power was perused at 415 nm. Catalase (CAT) and peroxidase (POD) were extricated by homogenization of the crisp leaf tissue in super cold refined water (1:10) with a chilled mortar and pestle. The response blend for CAT catalyst measured by the strategy for contained 0.005 M hydrogen peroxide in 0.025 mM potassium phosphate cushion (pH 7.0) and was institutionalized against 0.1 N KMnO₄. The response was begun by including 1 ml of the appropriately weakened compound concentrate. After 5 min, the response was halted by including 2 ml of 2 N H₂SO₄. Spaces, wherein sulfuric corrosive was added to the response blend before the option of the compound concentrate were run at the same time. Unused hydrogen peroxide in the response blend was titrated against 0.1N KMnO₄. The response blend for POD tested by the technique for which contained 2 ml of 0.1 M phosphate support (pH 6.0), 1 ml of 0.01% H₂O₂ and 1 ml of 0.5% p-phenylenediamine. The response was started by including 1 ml of the compound concentrate to the above blend and was permitted to continue for 5 min. The response was ceased by including 4N H₂SO₄. Spaces, wherein sulfuric corrosive was added preceding the option of the protein extricate, were run all the while. For examine of SOD, APX and GR new leaf tissue was ground in 50 mM potassium phosphate cradle (pH 7.0), containing EDTA (1 mM) and PVP (2%) The concentrates were centrifuged at 15000 g for 10 min, and the supernatant was measured for the chemical exercises. All out SOD was tested by checking the hindrance of photochemical decrease of nitroblue tetrazolium (NBT) in 3 ml response blend, which containing 50 mM phosphate support (pH 7.0), 0.13 mM methionine, 75 µM NBT, 2 µM riboflavin, 0.1 mM EDTA, and 25 µl of the protein concentrate and read at 560 nm. One unit of proteins is characterized as the measure of chemical causing half restraint of NBT decrease. For deciding APX action according to the strategy for, the response blend contained 50 mM potassium phosphate cushion (pH 7.0), 0.5 mM ascorbate, and 0.1 mM H₂O₂.

3. Oxidative stress

Oxidative pressure mirrors a lopsidedness between the foundational indication of receptive oxygen species and a natural framework's capacity to promptly detoxify the responsive intermediates or to fix the subsequent harm. Aggravations in the typical redox condition of cells can cause dangerous impacts through the generation of peroxides and free radicals that harm all parts of the cell, including proteins, lipids, and DNA. Oxidative worry from oxidative digestion causes base harm, just as strand breaks in DNA. Base harm is generally aberrant and brought about by responsive oxygen species (ROS) produced, for example O₂⁻ (superoxide radical), OH (hydroxyl radical) and H₂O₂ (hydrogen peroxide). [1] Further, some receptive oxidative species go about as cell emissaries in redox flagging. In this way,

oxidative pressure can cause disturbances in typical components of cell flagging.

4. Reactive Oxygen Species (ROS)

An unavoidable result of high-impact digestion is generation of responsive oxygen species (ROS). ROS incorporate free radicals, for example, superoxide anion (O₂⁻), hydroxyl radical (•OH), just as nonradical atoms like hydrogen peroxide (H₂O₂), singlet oxygen (1O₂, etc. Stepwise decrease of sub-atomic oxygen (O₂) by high-vitality introduction or electron-exchange responses prompts generation of the very receptive ROS. In plants, ROS are constantly shaped by the inescapable spillage of electrons onto O₂ from the electron transport exercises of chloroplasts, mitochondria, and plasma films or as a result of different metabolic pathways limited in various cell compartments. Ecological burdens, for example, dry season, saltiness, chilling, metal harmfulness, and UV-B radiation just as pathogens assault lead to upgraded age of ROS in plants because of interruption of cell homeostasis. All ROS are amazingly unsafe to living beings at high focuses. At the point when the dimension of ROS surpasses the safeguard instruments, a phone is said to be in a condition of "oxidative pressure." The improved creation of ROS amid natural burdens can represent a risk to cells by causing peroxidation of lipids, oxidation of proteins, harm to nucleic acids, compound hindrance, initiation of customized cell passing (PCD) pathway and eventually prompting demise of the cells.

Despite their destructive activity, ROS are well-described second messengers in a variety of cell forms including resistance to ecological burdens. Regardless of whether ROS will go about as harming or flagging atom relies upon the sensitive balance between ROS creation and rummaging. As a result of the multifunctional jobs of ROS, it is vital for the cells to control the dimension of ROS firmly to stay away from any oxidative damage and not to dispense with them totally. Rummaging or detoxification of overabundance ROS is accomplished by an effective antioxidative framework containing the nonenzymic just as enzymic cell reinforcements. The enzymic cell reinforcements incorporate superoxide dismutase (SOD), catalase (CAT), guaiacol peroxidase (GPX), proteins of ascorbate-glutathione (AsA-GSH) cycle, for example, ascorbate peroxidase (APX), monodehydroascorbate reductase (MDHAR), dehydroascorbate reductase (DHAR), and glutathione reductase (GR) [21]. Ascorbate (AsA), glutathione (GSH), carotenoids, tocopherols, and phenolics fill in as strong nonenzymic cancer prevention agents inside the cell. Different laborers have announced expanded exercises of numerous proteins of the cell reinforcement resistance framework in plants to battle oxidative pressure incited by different ecological anxieties. Support of a high cell reinforcement ability to rummage the poisonous ROS has been connected to expanded resistance of plants to these natural anxieties. Impressive advancement has been made in improving pressure instigated oxidative pressure resilience in yield plants by creating transgenic lines with modified dimensions of cell reinforcements. Concurrent articulation of numerous cell reinforcement compounds has been appeared to be more successful than single or twofold articulation for creating

transgenic plants with upgraded resistance to different natural burdens. The present audit centers around sorts of ROS, their site of creation, and their job as courier and inducer of oxidative pressure.

5. Sites of production

ROS are a gathering of free radicals, receptive atoms, and particles that are gotten from O₂. It has been evaluated that 1% of O₂ devoured by plants is occupied to create ROS [27] in different subcellular loci, for example, chloroplasts, mitochondria, peroxisomes. ROS are all around perceived for assuming a double job as both injurious and helpful species relying upon their fixation in plants. At high fixation ROS influence harm to biomolecules, though at low/moderate focus it goes about as second ambassador in intracellular flagging falls that intervene a few reactions in plant cells.

6. Effects of oxidative stress on the body

Oxidation is a typical and vital procedure that happens in your body. Oxidative worry, then again, happens when there's an irregularity between free extreme action and cancer prevention agent action. When working appropriately, free radicals can help fend off pathogens. Pathogens lead to contaminations.

At the point when there are sans more radicals present than can be kept in parity by cell reinforcements, the free radicals can begin doing harm to greasy tissue, DNA, and proteins in your body. Proteins, lipids, and DNA make up a

substantial piece of your body, with the goal that harm can prompt an immense number of infections after some time. These include:

- diabetes
- atherosclerosis, or the hardening of the blood vessels
- inflammatory conditions
- high blood pressure, which is also known as hypertension
- heart disease
- neurodegenerative diseases, such as Parkinson's and Alzheimer's
- cancer

7. Conclusion

ROS are unavoidable side-effects of ordinary cell digestion. ROS are produced by electron transport exercises of chloroplast, mitochondria, and plasma film or as a result of different metabolic pathways confined in various cell compartments. Under ordinary development condition, ROS generation in different cell compartments is low. Nonetheless, different natural anxieties, for example, dry spell, saltiness, chilling, metal danger, and UV-B, whenever delayed over to a limited degree, upset the cell homeostasis and improve the generation of ROS. ROS assume two different jobs in plants; in low fixations they go about as flagging particles that intercede a few plant reactions in plant cells, including reactions under anxieties, though in high focuses they cause worsening harm to cell segments.

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